Trends in the Molecular Epidemiology and Genetic Mechanisms of Transmitted Human Immunodeficiency Virus Type 1 Drug Resistance in a Large US Clinic Population

Soo-Yon Rhee,1 Dana Clutter,1 W. Jeffrey Fessel,2 Daniel Klein,1 Sally Slome,1 Benjamin A. Pinsky,3 Julia L. Marcus,4 Leo Hurley,7 Michael J. Silverberg,7 Sergei L. Kosakovsky Pond,9 and Robert W. Shafer1

1Division of Infectious Diseases, Department of Medicine, Stanford University; 2Department of Internal Medicine, Kaiser Permanente Northern California, San Francisco; 3Department of Infectious Diseases, Kaiser Permanente Northern California, San Leandro; 4Department of Infectious Diseases, Kaiser Permanente Northern California, Oakland, and 5Department of Pathology, Stanford University, California; 6Harvard Medical School and Harvard Pilgrim Health Care Institute, Boston, Massachusetts; 7Division of Research, Kaiser Permanente Northern California, Oakland; and 8Department of Biology, Temple University, Philadelphia, Pennsylvania

Background. There are few large studies of transmitted drug resistance (TDR) prevalence and the drug resistance mutations (DRMs) responsible for TDR in the United States.

Methods. Human immunodeficiency virus type 1 (HIV-1) reverse transcriptase (RT) and protease sequences were obtained from 4253 antiretroviral therapy (ART)–naive individuals in a California clinic population from 2003 to 2016. Phylogenetic analyses were performed to study linkages between TDR strains and selection pressure on TDR-associated DRMs.

Results. From 2003 to 2016, there was a significant increase in overall (odds ratio [OR], 1.05 per year [95% confidence interval (CI), 1.03–1.08]; P < .001) and nonnucleoside RT inhibitor (NNRTI)–associated TDR (OR, 1.11 per year [95% CI, 1.08–1.15]; P < .001). Between 2012 and 2016, TDR rates to any drug class ranged from 15.7% to 19.2%, and class–specific rates ranged from 10.0% to 12.8% for NNRTIs, 4.1% to 8.1% for RT inhibitors (NRTIs), and 3.6% to 5.2% for protease inhibitors. The thymidine analogue mutations, M184V/I and the tenofovir-associated DRMs K65R and K70E/Q/G/N/T accounted for 82.9%, 7.3%, 10.0% to 12.8% for NNRTIs, 4.1% to 8.1% for nucleoside RT inhibitors (NRTIs), and 3.6% to 5.2% for protease inhibitors. The thymidine analogue mutations, M184V/I and the tenofovir-associated DRMs K65R and K70E/Q/G/N/T accounted for 82.9%, 7.3%, 10.0% to 12.8% for NNRTIs, 4.1% to 8.1% for nucleoside RT inhibitors (NRTIs), and 3.6% to 5.2% for protease inhibitors. The thymidine analogue mutations, M184V/I and the tenofovir-associated DRMs K65R and K70E/Q/G/N/T accounted for 82.9%, 7.3%, 10.0% to 12.8% for NNRTIs, 4.1% to 8.1% for nucleoside RT inhibitors (NRTIs), and 3.6% to 5.2% for protease inhibitors.

Conclusions. Although TDR has increased significantly in this large cohort, many TDR strains are unlikely to influence the activity of currently preferred first-line ART regimens. The high proportion of DRMs associated with infrequently used regimens combined with the clustering of TDR strains suggest that some TDR strains are being transmitted between ART-naive individuals.

Keywords. HIV-1 transmission; HIV-1 drug resistance; mutation; reverse transcriptase; protease.
transcriptase (RT) and protease between January 2003 and December 2016 at the Stanford University Healthcare Diagnostic Virology Laboratory. KPNC is estimated provide care to approximately 25% of the insured population in Northern California [12]. Cohort individuals were characterized by age, gender, race, HIV acquisition risk factor, and baseline plasma HIV-1 RNA level and CD4 count. For ART-naive individuals having >1 resistance test, the virus sequence of the first test was analyzed. Sequences from 13 individuals not encompassing protease positions 10–90 and RT positions 40–240 were excluded. The Stanford University and KPNC institutional review boards approved this study.

Prevalence and Temporal Trend of TDR
TDR was defined as the presence of 1 or more mutations from the World Health Organization 2009 list of 93 surveillance DRMs (SDRMs) at 43 positions, including 34 nucleoside reverse transcriptase inhibitor (NRTI)–, 19 NNRTI–, and 40 protease inhibitor (PI)–associated DRMs [13]. We determined the proportion of individuals with TDR to each class and with multiclass TDR. We used generalized binomial logistic regression models to assess the relationship between sample year and TDR and calculated the odds ratio for yearly increases in TDR prevalence.

A subset of the NRTI-associated SDRMs were classified as thymidine analogue mutations (TAMs) including M41L, D67N/G, K70R, L210W, T215Y/F, K219Q/E/R/N, and the T215 revertants T215C/D/E/I/S/V (which evolve from T215F/Y in the absence of selective drug pressure). Several additional DRMs not on the SDRM list were analyzed including (1) the primarily tenofovir disoproxil fumarate (TDF)–selected DRMs A62V, K65N, and K70G/N/Q/S/T [14] and (2) the primarily rilpivirine (RPV)–selected DRMs E138A/G/K/Q, of which E138A is polymorphic, occurring in 1%–4% of viruses from ART-naive individuals [15, 16].

The Stanford HIV Drug Resistance Database (HIVDB) genotypic resistance interpretation program was used to quantify the clinical impact of DRMs on the NRTIs abacavir (ABC), TDF, zidovudine (ZDV), and the cytosine analogues lamivudine and emtricitabine (3TC/FTC); the NNRTIs efavirenz (EFV), RPV, and etravirine (ETR); and the pharmacologically boosted PIs lopinavir/ritonavir (LPV/r), atazanavir (bATV), and darunavir (bDRV) [17].

Phylogenetic Analyses
We performed 2 phylogenetic analyses to characterize TDR transmission dynamics. The first, which included solely ART-naive individuals, examined the extent of sequence similarity among TDR strains. The second, which included both ART-naive and -experienced individuals, determined the proportion of TDR strains that were similar to strains from ART-experienced individuals. For both phylogenetic analyses we concatenated the 297 protease nucleotides and the 900 nucleotides encompassing RT positions 1–300, masked ambiguous nucleotides and those encoding SDRMs, and used IQTREE to create a maximum likelihood tree using a generalized time-reversible substitution model with site-to-site rate variation modeled by the discrete gamma distribution with 4 rate classes and an invariant proportion (GTR + I + G4), and generated bootstrap support using 1000 replicates [18].

For the first analysis, we used the complete set of sequences from the ART-naive cohort. Following tree construction, we used ClusterPicker to identify sequence clusters defined as sequences of subtrees having a maximum pairwise distance between sequences <0.02 and a bootstrap value >70% [19]. We then determined the proportion of TDR viruses in a cluster containing other viruses sharing the same SDRMs.

For the second analysis, we combined the sequences from the ART-naive cohort with sequences from ART-experienced individuals in KPNC who underwent resistance testing at Stanford University Healthcare since such testing began in 1998. For this analysis, we used a higher pairwise distance threshold of 0.04, albeit with a stricter bootstrap value of ≥90. The higher distance threshold and stricter bootstrap value defined clades in which all leaves shared a common ancestor but, as a result of the higher distance threshold, were less likely to be closely related epidemiologically than individuals in the ART-naive clusters. We then determined the proportion of individuals with TDR that had viruses in the same clade as a virus sequence, with the same SDRMs, from an ART-experienced individual with acquired drug resistance.

For the largest TDR cluster, we performed Bayesian phylogenetic inference of time-measured trees using Markov chain Monte Carlo (MCMC) sampling implemented in BEAST (Bayesian Evolutionary Analysis Sampling Trees) version 1.8.4 software [20]. The substitution process was modeled according to the GTR + I + G4 model and an exponential growth model was specified as coalescent tree prior. Independent MCMC analyses were run for 10 million generations, sampling every 5000 generations, and the first 10% of the samples was discarded as burn-in before combining the samples. The runs were investigated based on effective sample size calculated using Tracer [21]. Then a maximum clade credibility tree was selected from the posterior tree distribution and visualized using FigTree [22].

To determine if viral lineages in ART-naive individuals underwent diversifying positive selection at SDRM sites, we applied the fixed effects likelihood test in HyPhy version 2.3.10. The test was restricted only to terminal branches leading to sequences from ART-naive individuals, or those internal branches whose descendants were all ART-naive, in the phylogenetic tree containing sequences from both ART-naive and -experienced individuals [23].

RESULTS
Study Cohort
Between January 2003 and December 2016, 4253 HIV-1–infected ART-naive individuals underwent genotypic
resistance testing. The median number of ART-naive individuals per year was 304 (range, 178–355). The median age was 39.5 and 90.5% were male (Table 1). The cohort composition was 42.5% white, 20.9% African American, 19.2% Latino, 8.0% Asian, 0.4% American Indian/Alaska Native, and 9.0% unknown. The HIV-1 acquisition risk factors included men who have sex with men (59.2%), heterosexual contact (18.1%), bisexual contact (10.4%), intravenous drug use (6.2%), receipt of transfusion (0.4%), and perinatal infection (0.1%). Acquisition risk factors were unrecorded for 5.6% of the population.

At the time of genotypic resistance testing, the median CD4 count was 349 cells/µL (interquartile range [IQR], 180–527 cells/µL) and the median plasma HIV-1 RNA level was 4.5 log copies/mL (IQR, 4.0–5.1 log copies/mL). One hundred seventy-nine (4.2%) individuals had repeated genotypic resistance tests before starting ART. Subtype B accounted for 95.3% of viruses. The remaining 4.7% included subtype C (1.3%), circulating recombinant form (CRF) 01_AE (1.2%), CRF02_AG (0.5%), A (0.4%), and other subtypes and CRFs (1.3%).

The 4253 sequences contained a median of 0.67% mixtures per nucleotide (IQR, 0.17%–1.33%). There was a yearly 1.06-fold (95% confidence interval [CI], 1.05–1.08; P < .001) increase in the odds of having a sequence with <0.5% mixtures, a proxy for recent infection [24–26], with 29.8% of 2003 and 52.3% of 2016 samples having <0.5% mixtures. Consistent with the concept that over time the cohort underwent resistance testing earlier during infection, there was also a significant yearly increase in CD4 cell count (regression coefficient of 10.9 cells/µL; P < .001). The mean CD4 count in 2003 was 259 cells/µL and in 2016 was 419 cells/µL.

### TDR Prevalence and Temporal Trends

Over the 14 year-study period, 591 of 4253 (13.9%) individuals had TDR. The overall proportions with NNRTI, NRTI, PI, and multiclass TDR were 7.2%, 5.8%, 3.2%, and 1.9%, respectively. Table 1 shows that subtype B and Asian race were associated with an increased TDR risk whereas African American race was associated with a reduced TDR risk. TDR risk was not significantly influenced by age, gender, acquisition risk factor, CD4 count, or plasma HIV-1 RNA level.

Figure 1 shows that there was a significant yearly increase in overall (odds ratio [OR], 1.05 [95% CI, 1.03–1.08]; P < .001) and NNRTI-associated TDR (OR, 1.11 [95% CI, 1.08–1.15]; P < .001). There was also weakly significant increases in PI-associated (OR, 1.05 [95% CI, 1.0–1.09]; P = .05) and multiclass TDR (OR, 1.09 [95% CI, 1.03–1.16]; P = .005). There was no significant temporal change in NRTI-associated TDR (OR, 0.99 [95% CI, .95–1.02]; P = .4). Supplementary Table 1 lists the proportions of individuals with TDR by drug class for each study year.

### SDRMs Responsible for TDR

Among viruses from the 591 individuals with TDR, there were a total of 960 SDRMs (1.62 per individual): 387 (65.5%) had 1 SDRM, 128 (21.7%) had 2 SDRMs, and 76 (12.8%) had ≥3 SDRMs. Figure 2 displays the temporal trends of the most common NNRTI, NRTI, and PI SDRMs. Supplementary Tables 2–4 contain the complete list of SDRMs and their proportions by ART class.

The mutations K103N/S, Y181C, Y188L, and G190A accounted for 88.5% of the 348 NNRTI SDRMs. The mutations E138A/G/K/Q, which are associated with reduced RPV susceptibility but are not on the SDRM list, occurred in 99 individuals, including 90 without an SDRM. The polymorphic mutation E138A accounted for 87 of these 99 mutations [15, 16]. For the entire population, the proportion with predicted low-, intermediate-, or high-level NNRTI resistance was 7.8% for EFV, 5.1% for RPV, and 1.6% for ETR.

### Table 1. Association of Study Cohort Characteristics With Transmitted Drug Resistance

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Overall (N = 4253)</th>
<th>Wild-type (n = 3862)</th>
<th>TDR (n = 591)</th>
<th>P Valuea</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, y, mean</td>
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<td>39.2</td>
<td>.4</td>
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<tr>
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<td>4.5</td>
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<tr>
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<td>90.3</td>
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<tr>
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<td></td>
<td></td>
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<td>0.5</td>
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<td>8.7</td>
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<td>18.1</td>
<td>18.3</td>
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<tr>
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<td>10.4</td>
<td>10.7</td>
<td>.91</td>
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<tr>
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<td>6.2</td>
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<tr>
<td>Transfusion</td>
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<td>.17</td>
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<td>.17</td>
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<tr>
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<td>0</td>
<td>.2</td>
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<tr>
<td>Other subtypes/CRFs</td>
<td>1.3</td>
<td>1.3</td>
<td>0.9</td>
<td>.4</td>
</tr>
</tbody>
</table>

Except for continuous data, given as means, data are shown as percentages of the total number of individuals indicated in the column headers.

Abbreviations: CRF, circulating recombinant form; HIV-1, human immunodeficiency virus type 1; MSM, men who have sex with men; TDR, transmitted drug resistance.

a Student t test was used for comparisons of continuous data (age, HIV-1 RNA, and CD4 cell count). The χ² test was used for comparisons of categorical data.

b Intravenous drug users included individuals with or without MSM as a risk factor.
Of the 409 NRTI SDRMs, 82.9% were TAMs (including T215 revertant mutations). However, the primary TAMs T215F/Y were present in just 5 individuals. M184V/I occurred in 20 individuals, comprising 7.3% of NRTI SDRMs. The non-TAMs K65R, L74V/I, Y115F, and Q151M each occurred in just 1 or 2 individuals. The TDF-associated SDRM K70E did not occur. K70N/T/S, which are not on the SDRM but are associated with low-level TDF resistance, occurred in 3 individuals including 1 without an SDRM. A62V, a non-SDRM TDF-associated accessory DRM, occurred in 10 individuals. For the entire population, the proportion with predicted low-, intermediate-, or high-level NRTI resistance was 4.9% for ZDV, 2.8% for ABC, 2.2% for TDF, and 0.7% for 3TC/FTC.

Of the 203 PI-associated SDRMs, the most common were L90M (33.5%), M46I/L (19.7%), I54V (10.8%), V82A/L/T (10.4%), and D30N plus N88D (9.8%). The next most common SDRMs, V32I, I50V/L, L76V, and I84V, each occurred in ≤5 individuals. For the entire population, the proportion with predicted low-, intermediate-, or high-level PI resistance was 2.2% for bATV, 2.1% for LPV/r, and 0.3% for bDRV.

![Figure 1](https://example.com/fig1.png)

**Figure 1.** Temporal trends in the yearly proportion of individuals with transmitted drug resistance, by drug class. The fitted lines show the effect of sample years in generalized binomial logistic regression models. Abbreviations: NNRTI, nonnucleoside reverse transcriptase inhibitor; NRTI, nucleoside reverse transcriptase inhibitor; PI, protease inhibitor; TDR, transmitted drug resistance.

![Figure 2](https://example.com/fig2.png)

**Figure 2.** Temporal trends in the yearly proportion of individuals with the most commonly detected nucleoside reverse transcriptase inhibitor, nonnucleoside reverse transcriptase inhibitor, and protease inhibitor surveillance drug resistance mutations. The fitted lines show the effect of sample years in generalized binomial logistic regression models. Abbreviations: DRM, drug resistance mutation; NNRTI, nonnucleoside reverse transcriptase inhibitor; NRTI, nucleoside reverse transcriptase inhibitor; PI, protease inhibitor; SDRM, surveillance drug resistance mutation; TAM, thymidine analogue mutation.
Virus Sequence Clustering

The median uncorrected intrasubtype pairwise distance between sequences in the ART-naive cohort was 0.055 (IQR, 0.047–0.062). Of these sequences, 43.8% (1864) were present in 1 of 669 clusters. Of these, 134 contained 1 or more sequences with an SDRM, comprising 47.0% of the 591 individuals with TDR. Figure 3A and 3B show the distribution of the number of clusters by cluster size for all 669 clusters and for the 134 clusters containing 1 or more sequences with an SDRM.

Of the 669 clusters, 97.3% comprised subtype B viruses; 2.7% (n = 18) comprised viruses belonging to other subtypes including C (6), CRF01_AE (6), CRF02_AG (2), A (1), D (1), F (1), and G (1). One CRF01_AE cluster contained 12 viruses without SDRMs. The remaining non–subtype B clusters contained 2–3 viruses including 2 clusters with SDRMs.

Composition of Clusters Containing TDR Viruses

Of the 134 clusters containing viruses with an SDRM, 82 contained ≥2 viruses sharing 1 or more SDRMs (Figure 4). Within these 82 clusters, the median cluster size was 2 (range, 2–12) and the median pairwise distance was 0.0064. The median range in years from the earliest to most recent sample was 4.0 years (IQR, 2.0–6.8 years). In 23.2% (n = 19) of the 82 TDR clusters, each virus was obtained within the same 12-month period.

Overall, 68 of 82 (82.9%) clusters in Figure 4 consisted entirely of viruses with SDRMs including 27 containing 64 NNRTI TDR viruses, 24 containing 71 NRTI TDR viruses, 7 containing 16 PI TDR viruses, and 10 containing 37 viruses with SDRMs conferring resistance to >1 drug class. The remaining 14 clusters contained 1 or more viruses without SDRMs in addition to viruses with an SDRM. Overall, 55 of the 82 (67.1%) clusters were homogeneous in that they contained identical SDRMs; 27 (32.9%) were heterogeneous containing 1 or more viruses without SDRMs or with different, albeit overlapping, SDRMs.

TDR Viruses Similar to Those From ART-experienced Individuals

The phylogenetic analysis of the sequences from the 4253 ART-naive individuals combined with 4064 sequences from 3155 ART-experienced individuals showed that 83 viruses from the 591 (14%) ART-naive individuals with TDR were in the same clade as a virus from an ART-experienced individual whose virus contained each of the SDRMs as the TDR virus. The median uncorrected genetic distance between these 83 virus pairs was 0.009 (IQR, 0.003–0.015).

Figure 5 contains a time-scaled Bayesian phylogenetic tree that illustrates in red the likely origin of the cluster of 12 viruses containing the NNRTI DRM Y181C plus the PI DRM L90M. The drug-resistant virus first emerged in 1999 in an individual who received sequential therapy with several drugs including ZDV, 3TC, stavudine, nevirapine, and saquinavir. This individual had 4 genotypes performed between 1998 and 2012, each of which is indicated in blue.

Positional Selection Pressure in Protease and RT

The fixed-effects likelihood test for selection pressure on the branches leading to the sequences from ART-naive individuals identified 29 codons in protease and RT that were subject to diversifying positive selection (P ≤ .01), but there was no evidence of positive selection at any of the 43 SDRM positions. Each of these 43 codons was inferred to be subject to purifying selection, and at 40 of the codons this finding was statistically significant (P ≤ .01, likelihood ratio test). Supplementary Table 5 summarizes the fixed-effects likelihood test results for all 43 SDRM positions.

DISCUSSION

In the 14 years since pretreatment HIV genotypic resistance testing became the standard of care in this Northern California clinic-based population, there has been a progressive increase in TDR. The increase occurred in multiple demographic groups and in individuals with diverse HIV-1 acquisition risk factors. However, because most TDR strains had NNRTI-associated DRMs or NRTI-associated TAMs, they are unlikely to influence...
Figure 4. Composition of surveillance drug resistance mutation (SDRM) patterns of the 82 clusters containing ≥2 viruses sharing ≥1 SDRM. Clusters were defined as including viruses with a maximum pairwise distance ≤0.02 and bootstrap support value ≥70%. Overall, 68 (82.9%) of the clusters consisted entirely of viruses with SDRMs and 55 (67.1%) were homogeneous (ie, containing the same SDRMs). Abbreviations: NNRTI, nonnucleoside reverse transcriptase inhibitor; NRTI, nucleoside reverse transcriptase inhibitor, NRTITAM, nucleoside reverse transcriptase inhibitor thymidine analogue mutation; PI, protease inhibitor; WT, wild-type.
the activity of the preferred first-line ART regimens, which infrequently include NNRTIs and very rarely include thymidine analogues. Our finding of an ongoing TDR increase is consistent with the high TDR levels observed in other North American studies [4–6, 8, 27]. The finding that NNRTI-associated TDR has been increasing is consistent with the global TDR trends [28, 29].

Consistent with a previous meta-analysis, a small subset of NNRTI-associated SDRMs accounted for the vast majority of NNRTI TDR [29]. Although the spectrum of NRTI-associated SDRMs was diverse, >80% of NRTI TDR cases were caused by TAMs, of which those other than T215Y/F may have little impact on currently used NRTIs [30]. The rarity of K65R,

Figure 5. Time-scale analysis of the large virus cluster containing viruses with the nonnucleoside reverse transcriptase inhibitor resistance mutation Y181C and the protease inhibitor resistance mutation L90M. Viruses were labeled with a randomly generated person identifier (PID), the sample year and month in four digits (two-digit sample year and two-digit month), and the viruses’ list of surveillance drug resistance mutations delimited by “_”. The values at the nodes represent posterior support values for the clusters obtained using Markov chain Monte Carlo sampling implemented in BEAST version 1.8.4 software. Leaf nodes for sequences from antiretroviral therapy (ART)–naive individuals are unfilled circles whereas those from ART-experienced individuals are filled circles. Viruses from 16 ART-naive individuals with Y181C + L90M are indicated in red, and 4 viruses with the same mutations from an ART-experienced individual are indicated in blue (PID 52). This ART-experienced individual appeared to acquire Y181C + L90M plus K70R in 1999 (sequence followed by an asterisk). One individual (PID 50) was also primarily infected with this virus as a 2002 sequence obtained prior to therapy contained Y181C + L90M (indicated in red with an open triangle) and later developed K103N (indicated in black with a closed triangle) after receiving multiple nucleoside reverse transcriptase inhibitor regimens followed by tenofovir/emtricitabine/efavirenz in combination with atazanavir/ritonavir and then raltegravir. There were an additional 3 individuals whose viruses likely originated from this virus strain but were not in the same cluster because their sequences differed from the 12 clustered viruses by >2.0%.
other less common TDF-associated mutations, and the primary TAMs T215Y/F indicates that transmitted TDF resistance is unusual. Moreover, the evolution of US treatment guidelines toward first-line regimens that include an integrase strand transfer inhibitor (INSTI) or bDRV means that the preferred first-line regimens are highly active in most patients with TDR [31].

The prevalence of TDR in chronically infected individuals is generally lower than in acutely infected individuals because in the years following initial infection, the least fit DRMs revert to wild type [32–35]. Indeed, our selection analyses shows that within the combined phylogeny of sequences from ART-naive and ART-experienced individuals, natural selection maintained existing amino acids at the SDRM sites along the ART-naive branches. Thus, despite their ability to persist in ART-naive populations, the overall trend for many SDRMs is toward a gradual reversion to wild type. Some of the increase in TDR prevalence may also have resulted from the trend to perform genotypic resistance testing earlier in infection as evidenced by the cohort’s progressively lower proportion of sequence mixtures and higher CD4 counts.

Endemic TDR strains emanating from a single instance of ART-selection pressure that spread among ART-naive individuals have different implications from TDR strains emanating from ART-experienced individuals. Natural selection maintained existing DRMs would be expected to filter out minority variants that may have originally been transmitted from an ART-experienced individual [36, 37].

The high proportion of DRMs associated with infrequently used ART regimens combined with the clustering of many TDR strains suggests that some proportion of TDR strains represent established drug-resistant lineages transmitted among ART-naive individuals. This hypothesis is consistent with findings from other countries with mature HIV-1 epidemics such as Switzerland and the United Kingdom, where it has been estimated that most TDR cases are transmitted from ART-naive individuals [34, 36, 38, 39]. Our dataset, however, is limited by the fact that KPNC provides care to about 25% of the insured population in Northern California [12], making it likely that many transmission events occurred with individuals not within our cohort and not captured in our phylogenetic analyses.

In conclusion, this is one of the largest studies of the trends and mutation patterns associated with HIV-1 TDR in the United States. The finding that a large proportion of TDR strains contain DRMs associated with regimens now used infrequently is consistent with the concept that many TDR cases are transmitted between ART-naive individuals. Although the frequency of transmitted INSTI resistance has been extremely rare in the United States [40], ongoing surveillance is required as INSTIs have been increasingly used for both first-line and salvage ART.

**Supplementary Data**

Supplementary materials are available at *Clinical Infectious Diseases* online. Consisting of data provided by the authors to benefit the reader, the posted materials are not copyrighted and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

**Notes**

**Acknowledgments.** The complete set of sequence data from 4253 antiretroviral therapy (ART-naive) individuals has been submitted to GenBank. Of the 4253 ART-naive individuals, sequences from 1144 individuals had been previously deposited and are available in GenBank and their accession numbers are listed in the Supplementary Materials. The sequences from the remaining individuals have been submitted to GenBank (MG941022–MG944215).

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**References**


21. Rambaut A, Drummond AJ. Tracer: MCMC trace analysis tool. 1.5 e.


